

Effect of Short-term Sulfur dioxide Fumigation on Photosynthesis in Sunflower and Wheat

Adeola Olateju Grillo

Abstract

Wheat and sunflower plants were exposed to 0.289ppm, 0.578ppm, 1.156ppm, and 2.312ppm of SO₂ for one hour. Photosynthetic rates were measured, after treatment, with a LI-COR infra-red gas analyzer, model 6200. Photosynthetic stimulation was found to occur in wheat at 0.289ppm and 0.578ppm, photosynthetic rates were about the same as the control at 1.156ppm, while inhibition occurred at 2.312ppm. Photosynthetic inhibition occurred in sunflower at all fumigation concentrations.

Introduction

Short-term fumigation with SO₂, often performed with plants grown in protected environments, does not necessarily reflect conditions in the open field. However, there are advantages in investigating its effects. First, it simulates a common situation in urban areas where SO₂ peaks may reach several hundred parts per billion. Second, it can be used in simple mechanistic studies of interactions between photosynthesis, the stomatal mechanism, and uptake, since these are fundamentally the same with chronic exposures. Third, the effects of short-term fumigation can be used as diagnostic tools, when these effects have been documented to reflect a situation in the "real world" (Saxe, 1991).

Generally, the effect of fumigation on photosynthesis depends on the pollutant, pollutant concentration, and exposure times. It also depends on the intake rate of the plant and this varies with species, cultivar, age, and environmental conditions such as temperature, humidity, light, and CO₂ concentration.

Photosynthetic inhibition was normally found in short-term fumigation when the concentration multiplied by the duration was above 400 ppb h (parts per billion hour), and at the lower concentrations only after 2h or more of exposure. There have been exceptions where inhibition was found at lower exposure concentrations and durations, while a few other authors have found low exposure levels to stimulate photosynthesis. At higher concentrations and durations, however, the stimulation always turned to inhibition (see Table 1 for examples).

The aim of this research was to study and compare the effect of short-term SO₂ fumigation on photosynthesis in wheat and sunflower plants, using concentrations above and below 400 ppb h.

Materials and Methods

Pots containing three or four plants were used for the experiments. Four to seven week old plants were placed, one pot at a time, in an 18.8L polycarbonate

plastic chamber and injected with SO₂ ranging from 0.289 ppb to 2.312 ppb. The pots were removed after 1h.

Table 1. Changes in net photosynthesis due to short-term fumigation of SO₂.

Reference	Species and Cultivar	[SO ₂] (ppb)	Duration	PS-response (% control)
INHIBITION				
Carlson	soybean	450	2h	73
Furukawa et al.	sunflower	1500	30 min	27
Matsuoka	rice	170	5h	95
Black and Unsworth	broad bean	35	2h	85
STIMULATION FOLLOWED BY INHIBITION				
Takemoto and Noble	soybean	250	1h	111
	strain T219	500	1h	91
		500	2h	72
Muller et al.	soybean	79	4h	137
		405	3h 40 min	83
Winner and Mooney	saltbush	500	2h	110
		500	7h	100

conc, concentration; ppb, parts per billion; cv, cultivar.

Photosynthetic rates were measured with a LI-COR IRGA (infra-red gas analyzer), model 6200. The analyzer was set to make three ten-second observations per leaf. One leaf was chosen per plant and enclosed in a 1L leaf chamber. The leaf area enclosed was measured by cutting out the area on a

sheet of paper and weighing. A known area of the same kind of paper was weighed, and leaf area calculated by using a conversion factor (known area*weight of cut paper/known weight). The measured leaf areas were used by the analyzer to calculate raw photosynthetic rates ($\mu\text{mol m}^{-2} \text{s}^{-1}$). Each raw rate of photosynthesis was then divided by the light intensity ($\mu\text{mol m}^{-2} \text{s}^{-1}$) during each measurement.

Statistical analyses were performed using one-way ANOVA to detect significant differences between fumigation concentrations and the control, and within fumigation concentrations.

Results

Photosynthesis was inhibited in the sunflower leaves by SO_2 fumigation. The effects due to the fumigation concentrations differed significantly with the control at the 0.05 level of significance. However, there were no significant differences within the fumigation concentrations. (Table 2).

The effect on wheat was stimulation followed by inhibition. Stimulation occurred in the first two SO_2 concentrations. Photosynthetic rate in the third concentration was about the same as the control, while it was inhibited in the highest concentration (Table 3). Statistical analysis at the 0.05 level of significance showed that the effect due to fumigation at the highest concentration produced a significant difference in photosynthesis compared to the other fumigation concentrations and the control. There was also a significant difference in effects due to fumigation of 0.289 ppm compared to the control, and between 0.289 ppm and 1.156 ppm. However there were no significant differences between 0.289 ppm and 0.578 ppm, nor between the control and 1.156 ppm.

Table 2. Mean and percentage photosynthesis of sunflower plants.

# of samples	SO_2 conc (ppm)	Mean photo	SD photo	% photo (expressed as % of control)
12	control	0.0755	0.1078	100.00
8	0.2890	0.0111	0.0054	15.52
8	0.5780	0.0140	0.0031	18.58
8	1.1560	0.0132	0.0050	17.55
8	2.3120	0.0059	0.0026	7.82

Discussion

Short-term exposures to SO_2 may cause photosynthesis to decline (or increase) both in response to direct effects on chloroplast structure and function, and indirectly in response to effects on

stomata and other factors with a secondary influence on photosynthesis (Saxe, 1991).

Table 3. Mean and percentage photosynthesis of wheat plants.

# of leaf samples	SO_2 conc (ppm)	Mean photo	SD photo	% photo (expressed as % of control)
20	control	0.04	0.02	100.00
8	0.2890	0.06	0.05	150.00
12	0.5780	0.05	0.03	125.00
8	1.1560	0.04	0.02	100.00
8	2.3120	0.01	0.02	50.00

Effect on Chloroplasts

Structure. Swelling of the spaces within the thylakoids, reduction or disappearance of grana, gradual changes of chloroplast shape from ellipsoidal to oval to spherical, and invaginations from the inner membrane of the chloroplast membrane occur with SO_2 fumigation. Granulation of stroma also occurs at later stages of injury (Treshow and Anderson, 1991).

Function. Ribulose-1,5-bis-phosphate-carboxylase (RuBPC) is the enzyme that catalyzes the addition of CO_2 to ribulose 1,5- bisphosphate to form two molecules of 3-phosphoglycerate (carbon fixation). Ziegler (1972) demonstrated that sulphite can inhibit RuBPC *in vitro* by competing with HCO_3^- , enough to explain *in vitro* inhibition of photosynthesis by sulphite.

Other studies (Sugahara, 1984, Shimazaki, 1988) show that high levels of SO_2 and other gaseous pollutants induce effects on the thylakoid electron transport processes. It has even been found that low SO_2 concentrations induced a stimulation of electron transport in cereals, in accordance with the increase sometimes observed in photosynthesis with low pollutant exposures (Saxe, unpublished data). This would explain why the wheat plants were not inhibited until the highest concentration of fumigation.

The few studies on photorespiration are conflicting. Increases of photorespiration with increasing SO_2 concentrations have been shown that may result from the greater use of energy in repair or replacement processes (Kozial and Jordan, 1978). On the other hand, SO_2 may reduce or arrest rates of photorespiration (Ziegler, 1975). Since such inhibition would enhance net photosynthesis, it would be beneficial to the plant.

Another mechanism of action for high SO_2 concentrations is in causing the degradation of chlorophylls to phaeophytins, leading to early senescence (LeBlanc and Rao, 1975).

Effect on stomata. Hunt and Black (1988), Winner and Mooney (1980b) and several others, showed for a number of species that inhibition of photosynthesis by

acute SO₂ exposures could at the most be partly explained by stomatal closure. Previous studies (Takemoto and Noble, 1982; Biggs and Davis, 1980) show that the stomatal response to low doses is initial opening, but this rarely stimulates photosynthesis. However at 500-600ppb h and with exposures of two hours or longer, the stomata typically closes, which protects plants against further uptake of injurious SO₂.

The opening of stomata in field bean, *Vicia faba*, by low SO₂ concentrations was suggested to be caused by structural injury to surrounding epidermal cells before injury to the guard cells themselves (Black and Black, 1979); once guard cells were also injured, stomatal closure followed. In addition, Bonte and Cormis (1979) claimed acute SO₂ to have a direct effect on the stomatal metabolism in *Pelargonium*, concluding that SO₂ must affect the energy-requiring (opening and closing) mechanism directly.

Other factors. Increased CO₂ levels has been found to protect photosynthesis against SO₂ (Furukawa et al. 1979b). Majernik and Mansfield (1972) found that SO₂ stimulated opening of stomata at low CO₂ levels, while it tended to close stomata at high CO₂ levels in *Vicia faba*. They also found that at low light, 700ppb SO₂ stimulated stomatal opening as much as a doubling of the light intensity. Bonte and Louget (1975) found (*Pelargonium* plants) to be more injured in humid air, since this slowed the stomatal closing response to SO₂. Rist and Davis (1979) found that high temperature and humidity increased absolute stomatal conductance both with or without SO₂. Since it is this absolute conductance that determines uptake, leaf injury was far more severe at both high temperatures and high relative humidities.

Differences in response between wheat and sunflower could be due to species sensitivity, since it took a higher concentration of SO₂ to inhibit wheat, compared to sunflower. Environmental conditions were not kept constant during the experiments, which could have affected the rates of photosynthesis at the different concentrations. However, the effects of sulfur dioxide fumigation are quite clear with respect to inhibition in sunflower and stimulation followed by inhibition in wheat plants

Literature Cited

- Alscher, R. (1984). Effects of SO₂ on light modulated enzyme reactions. In "Gaseous Air Pollutants and Plant Metabolism" (M.J. Koziol and F.R. Whatley, eds). Chapter 14. Butterworths, London.
- Barton, J.R., McLaughlin, S.B. and McConahy, R.K. (1980). The effects of SO₂ on components of leaf resistance to gas exchange. Environmental Pollution (Ser. A) 21, 255-256.
- Biggs, A.R. and Davis D.D. (1980). Stomatal response to three birch species exposed to varying acute doses of SO₂. Journal of the American Society for Horticultural Science 105, 514-516.
- Black, C.R. and Black, V.J. (1979). The effects of low concentrations of sulphur dioxide on stomatal conductance and epidermal cell survival in field bean (*Vicia faba* L.). Journal of Experimental Botany 30, 291-298.
- Black, V.J. and Unsworth, M.H. (1979). Effects of low concentrations of sulfur dioxide on net photosynthesis and dark respiration of *Vicia faba*. Journal of Experimental Botany 30, 473-483.
- Bonte, J. and Cormis, L. de (1977). Inhibition, en anaerobiose, de la reaction de fermeture des stomates du *Pelargonium* en presence de dioxyde de soufre. Environmental Pollution 12, 125-133.
- Bonte, J. and Louguet, P. (1975). Interrelations entre la pollution par le dixoyde de soufre et le mouvement des stomates chez le *Pelargonium x hortum*: effets de l'humidite relative et de la teneur en gaz carbonique de l'air. Physiologie Vegetale 13, 527-537.
- Carlson, R.W. (1983). The effects of SO₂ on photosynthesis and leaf resistance at varying concentrations of CO₂. Environmental Pollution (Ser A)32, 11-38.
- Furukawa, A., Koike, A., Hozumi, K. and Tutsuka, T. (1979). The effects of SO₂ on photosynthesis in poplar leaves at various CO₂ concentrations. Journal of the Japanese Forestry Society 61, 351-356.
- Furukawa, A., Natori, T. and Totsuka, T. (1980). The effects of SO₂ on net photosynthesis in sunflower leaf. Research Report from the National Institute for Environmental Studies, Japan 65, 89-98.
- Hunt, G.A. and Black, V.J. (1988). Environmental stress: amplifier of physiological responses to SO₂ in plants. Environmental Pollution 53, 433-435.
- Koziol, M.J. and Jordan, C.E. (1978). Changes in carbohydrate levels in red kidney bean (*Phaseolus vulgaris* L.) exposed to sulphur dioxide. Journal of Experimental Botany 29, 1037- 1043.
- LeBlanc, D.C. and Rao, D.N. (1975). Effects of air

- pollutants on lichens and bryophytes. In "Responses of Plants to Air Pollution." (J.B. Mudd and T.T. Kozlowski, eds) 237-272, Academic Press, New York.
- Majernik, O. and Mansfield, T.A. (1972). Stomatal responses to raised atmospheric CO₂ concentration during exposure of plants to SO₂ pollution. *Environmental Pollution* 3, 1-7.
- Matsuoka, Y. (1978). Experimental studies of sulphur dioxide injury on rice plants and its mechanism. *Special Bulletin of the Chiba-Ken Agricultural Experimental Station* 7, 257-265.
- Muller, R.N., Miller, J.E. and Sprugel, D.G. (1979). Photosynthetic response of field-grown soybeans to fumigations with sulphur dioxide. *Journal of Applied Ecology* 16, 567-576.
- Rist, D.L. and Davis, D.D. (1979). The influence of exposure temperature and relative humidity on the response of pinto bean foliage to sulphur dioxide. *The American Phytopathological Society* 69, 231-235.
- Saxe, H. (1991). Photosynthesis and stomatal responses to polluted air, and the use of physiological and biochemical responses for early detection and diagnostic tools. *Advances in Botanical Research* 18, 1-128.
- Shimazaki, K.-I. (1988). Thylakoid membrane reactions to air pollutants. In "Air Pollution and Plant Metabolism". (S. Schulte-Hostede, N.M. Darrall, L.W. Blank and A.R. Wellburn, eds), pp 116-133. Elsevier Applied Science, London.
- Sisson, W.B., Booth, J.A. and Throneberry, G.O. (1981). Absorption of SO₂ by pecan (*Carya illinoensis* (Wang) K. Koch) and alfalfa (*Medicago sativa* L.) and its effect on net photosynthesis. *Journal of Experimental Botany* 32, 523-532.
- Sugahara, K. (1984). Effects of air pollutants on light reactions in chloroplast. In "Gaseous Air Pollutants and Plant Metabolism" (M.J. Koziol and F.R. Whatley, eds), Chapter 13. Butterworths, London.
- Takemoto, K. and Noble, R.D. (1982). The effect of short-term SO₂ fumigation on photosynthesis and respiration in soybean *Glycine max*. *Environmental Pollution (Series A)* 28, 67-74.
- Treshow, M. and Anderson, F.R. (1991). *Plant Stress from Air Pollution*. John Wiley and Sons, New York.
- Winner, W.E. and Mooney, H.A. (1980a). Ecology of SO₂ resistance: II. Photosynthetic changes of shrubs in relation to SO₂ absorption and stomatal behavior. *Oecologia* 44, 296-392.
- Winner, W.E. and Mooney, H.A. (1980b). Ecology of SO₂ resistance: III. Metabolic changes of C₃ and C₄ *Atriplex* species due to SO₂ fumigations. *Oecologia* 46, 49-54.
- Ziegler, I. (1972). The effects of SO₃ on the activity of ribulose-1,5-diphosphate carboxylase in isolated spinach chloroplasts. *Planta* 103, 155-163.
- Ziegler, I. (1975). The effect of SO₂ pollution on plant metabolism. *Residue Rev.* 56, 79-105.

Acknowledgements

I would like to thank Dr. Frye for all his help and encouragement.